



## Article

# Synthesis of Novel Benzenesulfonamide-Bearing Functionalized Imidazole Derivatives as Novel Candidates Targeting Multidrug-Resistant *Mycobacterium abscessus* Complex

Benas Balandis <sup>1</sup>, Povilas Kavaliauskas <sup>1,2,3,4,5,\*</sup> , Birutė Grybaitė <sup>1</sup>, Vidmantas Petraitis <sup>2,4,5</sup> , Rūta Petraitenė <sup>2,4</sup> , Ethan Naing <sup>2</sup>, Andrew Garcia <sup>2</sup>, Ramunė Grigalevičiūtė <sup>5</sup> and Vytautas Mickevičius <sup>1</sup>

<sup>1</sup> Department of Organic Chemistry, Kaunas University of Technology, Radvilėnų Rd. 19, LT-50254 Kaunas, Lithuania; benas.balandis@ktu.lt (B.B.); birute.grybaite@ktu.lt (B.G.); vytautas.mickevicius@ktu.lt (V.M.)

<sup>2</sup> Transplantation-Oncology Infectious Diseases Program, Division of Infectious Diseases, Department of Medicine, Weill Cornell Medicine of Cornell University, 1300 York Ave., New York, NY 10065, USA

<sup>3</sup> Institute for Genome Sciences, School of Medicine, University of Maryland Baltimore, 655 W. Baltimore Street, Baltimore, MD 21201, USA

<sup>4</sup> Institute of Infectious Diseases and Pathogenic Microbiology, Birštono Str. 38A, LT-59116 Prienai, Lithuania

<sup>5</sup> Biological Research Center, Lithuanian University of Health Sciences, Tilžės Str. 18/7, LT-47181 Kaunas, Lithuania

\* Correspondence: pok4001@med.cornell.edu



**Citation:** Balandis, B.; Kavaliauskas, P.; Grybaitė, B.; Petraitis, V.; Petraitenė, R.; Naing, E.; Garcia, A.; Grigalevičiūtė, R.; Mickevičius, V. Synthesis of Novel Benzenesulfonamide-Bearing Functionalized Imidazole Derivatives as Novel Candidates Targeting Multidrug-Resistant *Mycobacterium abscessus* Complex. *Microorganisms* **2023**, *11*, 935. <https://doi.org/10.3390/microorganisms11040935>

Academic Editor: Alexandra Tabaran

Received: 4 March 2023

Revised: 27 March 2023

Accepted: 1 April 2023

Published: 3 April 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

**Abstract:** Infections caused by drug-resistant (DR) *Mycobacterium abscessus* (*M. abscessus*) complex (MAC) are an important public health concern, particularly when affecting individuals with various immunodeficiencies or chronic pulmonary diseases. Rapidly growing antimicrobial resistance among MAC urges us to develop novel antimicrobial candidates for future optimization. Therefore, we have designed and synthesized benzenesulfonamide-bearing functionalized imidazole or S-alkylated derivatives and evaluated their antimicrobial activity using multidrug-resistant *M. abscessus* strains and compared their antimycobacterial activity using *M. bovis* BCG and *M. tuberculosis* H37Ra. Benzenesulfonamide-bearing imidazole-2-thiol compound **13**, containing 4-CF<sub>3</sub> substituent in benzene ring, showed strong antimicrobial activity against the tested mycobacterial strains and was more active than some antibiotics used as a reference. Furthermore, an imidazole-bearing 4-F substituent and S-methyl group demonstrated good antimicrobial activity against *M. abscessus* complex strains, as well as *M. bovis* BCG and *M. tuberculosis* H37Ra. In summary, these results demonstrated that novel benzenesulfonamide derivatives, bearing substituted imidazoles, could be further explored as potential candidates for the further hit-to-lead optimization of novel antimycobacterial compounds.

**Keywords:** benzenesulfonamides; imidazoles; S-alkylated; antimycobacterial activity; antifungal activity

## 1. Introduction

Infections caused by nontuberculous mycobacteria (NTM) remain a challenging and emerging public health threat, particularly in individuals undergoing chemotherapy or patients with underlying lung conditions [1]. The incidence of infections caused by NTM is increasing globally, and it can be challenging to diagnose and treat due to the diverse range of NTM species and varying patterns of antibiotic susceptibility. In addition, some NTM developed resistance to multiple antibiotics, making the infections caused by NTM difficult to treat and worsening the treatment prognosis [2–4]. Therefore, it is critical to develop novel small molecule antimicrobial candidates targeting NTM in particularly multidrug-resistant (MDR) strains.

*Mycobacterium abscessus* (*M. abscessus*) complex (MABC) is responsible for the majority of MDR NTM infections. MABC, consisting of genetically and phylogenetically related subspecies *M. abscessus* subsp. *abscessus*, *massiliense*, and *bolletii*, which often harbor multiple instinctive and acquired antimicrobial resistance determinants, as well as numerous virulence factors, make *M. abscessus* a clinically important pathogen [5]. Treatment or management of infections caused by *M. abscessus* requires long-term-to-lifetime treatment using multiple antibiotics. The treatment regime and duration is highly dependent on susceptibility of *M. abscessus* to macrolides [6]. Macrolide-susceptible *M. abscessus* infections require treatment using combinations of parenteral and inhalable antibiotics, which follows a maintenance period with at least three different inhalable and/or systemic antibiotics [7]. Macrolide-resistant MABC infections require a prolonged treatment duration, as well as an increased number of antibiotics, consequently making these infections extremely challenging. Therefore, it is crucial to discover novel candidates targeting MDR NTM with increased focus on *M. abscessus* [8].

Benzenesulfonamide scaffold have been widely used as a potent pharmacophore in medicinal chemistry [9,10]. Benzenesulfonamide nucleus-containing derivatives are widely explored in medicinal chemistry due to their ability to modulate various biological targets [11–16]. The sulfonamide group in the scaffold can act as a hydrogen bond acceptor and donor, providing a versatile platform for molecular modifications and improving the pharmacokinetic and pharmacodynamic properties of the targeted molecule. The benzene ring, on the other hand, is hydrophobic and contributes to the lipophilicity of the molecule. This can enhance the biologically active compound's ability to penetrate cell membranes or lipid layers that are found on the cell wall of various mycobacteria. Moreover, benzenesulfonamide derivatives have been previously reported to be good inhibitors of carbonic anhydrases (CAs) [17–19] and therefore exhibiting antimicrobial or anticancer activity. Moreover, CAs in *M. abscessus* have been previously reported to be a promising antimicrobial target since the pharmacological or molecular inhibition of CAs in *M. abscessus* leads to defective growth or virulence [20–23]. Therefore, benzenesulfonamide nucleus-bearing compounds could be attractive scaffolds for antimicrobial discovery targeting NTM.

Imidazole is a common heterocyclic scaffold that is often found in many natural and synthetic bioactive compounds and approved drugs. Molecules containing imidazole moiety can possess a wide variety of biological properties [24–31]. Structurally, imidazole can be further chemically modified with numerous substituents, making imidazole an extremely versatile scaffold [32–34]. Moreover, substituting a hydrophobic group at position 2 of the imidazole ring can improve the compound's lipophilicity, which can increase its membrane permeability and enhance the activity of antimicrobial compounds bearing an imidazole nucleus against mycobacteria.

In this study, we have synthesized novel benzenesulfonamide moiety-bearing functionalized imidazole derivatives containing various S-alkyl substituents and evaluated their antimicrobial activity against drug-resistant *M. abscessus* complex strains.

## 2. Materials and Methods

### 2.1. Reagents and Equipment Used for Synthesis and Characterization of Compounds

Reagents were purchased from Sigma-Aldrich (St. Louis, MO, USA). The melting points were determined on a MEL-TEMP (Electrothermal, Bibby Scientific Company, Burlington, NJ, USA) melting point apparatus and were uncorrected. IR spectra ( $\nu$ ,  $\text{cm}^{-1}$ ) were recorded on a Perkin–Elmer Spectrum BX FT-IR spectrometer using KBr pellets. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded in  $\text{DMSO-}d_6$  medium on Bruker Avance III (400, 101 MHz) spectrometer. Chemical shifts ( $\delta$ ) are reported in parts per million (ppm) calibrated from TMS (0 ppm) as an internal standard for  $^1\text{H}$  NMR, and  $\text{DMSO-}d_6$  (39.43 ppm) for  $^{13}\text{C}$  NMR. Elemental analysis was performed on a CE-440 elemental analyzer (Exeter Analytical Inc., North Chelmsford, MA, USA). The reaction course and purity of the synthesized compounds were monitored by TLC using aluminium plates precoated with silica gel 60 F<sub>254</sub> (MerckKGaA, Darmstadt, Germany).

## 2.2. Synthesis

Complete synthesis of compounds **2**, **4**, **5**, **7**, and **8**, as well as compounds **9**, **11**, **12**, **14**, **15**, **18b** was described in our previous study [35] and were resynthesized accordingly in this study. These compounds were further used for S-alkylation reactions. All of spectra data on compounds **2**, **4**, **5**, **7**, and **8** as well as compounds **9**, **11**, **12**, **14**, **15**, **18b** was described in our previous publication [35].

### General procedure for the synthesis of compounds **3** and **6**.

Amine **1** (1.72 g, 10 mmol) was dissolved in boiling water (40 mL). Then, the solution of corresponding  $\alpha$ -haloketone (12 mmol) in 10 mL of 1,4-dioxane was added dropwise to the mixture. The reaction mixture was heated at reflux for 2 h, then it was cooled down and the precipitate was filtered off, washed with diethyl ether, and recrystallized from 1,4-dioxane to afford compounds **3** and **6**.

**3-((2-(4-bromophenyl)-2-oxoethyl)amino)benzenesulfonamide (3)**. White solid, yield 3.12 g (85%); m.p. 236–237 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3382, 3262, 1692;  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 4.72 (s, 2H,  $\text{CH}_2$ ), 6.40 (br. s, 1H, NH), 6.84 (dd, 1H,  $J = 8.1$ , 2.3,  $H_{\text{ar}}$ ), 7.01 (d, 1H,  $J = 7.6$ ,  $H_{\text{ar}}$ ), 7.11 (br. s, 1H,  $H_{\text{ar}}$ ), 7.18 (br. s, 2H,  $\text{NH}_2$ ), 7.23 (t, 1H,  $J = 7.9$  Hz,  $H_{\text{ar}}$ ), 7.79 (d, 2H,  $J = 8.1$  Hz,  $H_{\text{ar}}$ ), 8.00 (d, 2H,  $J = 8.1$  Hz,  $H_{\text{ar}}$ );  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 49.96, 109.00, 112.96, 115.28, 127.68, 129.23, 129.93, 131.87, 134.04, 144.75, 148.49, 195.66; Anal. Calcd. for  $\text{C}_{14}\text{H}_{13}\text{BrN}_2\text{O}_3\text{S}$ : C 45.54; H 3.55; N 7.59 %. Found: C 45.57; H 3.55; N 7.54 %.

**3-((2-oxo-2-(4-(trifluoromethyl)phenyl)ethyl)amino)benzenesulfonamide (6)**. White solid, yield 2.97 g (83%); m.p. 228–229 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3390, 3268, 1698;  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 4.81 (s, 2H,  $\text{CH}_2$ ), 6.44 (br. s, 1H, NH), 6.84 (d, 1H,  $J = 7.7$ ,  $H_{\text{ar}}$ ), 7.03 (d, 1H,  $J = 7.7$ ,  $H_{\text{ar}}$ ), 7.13 (br. s, 1H,  $H_{\text{ar}}$ ), 7.19 (br. s, 2H,  $\text{NH}_2$ ), 7.24 (t, 1H,  $J = 7.9$  Hz,  $H_{\text{ar}}$ ), 7.95 (d, 2H,  $J = 8.0$  Hz,  $H_{\text{ar}}$ ), 8.26 (d, 2H,  $J = 8.0$  Hz,  $H_{\text{ar}}$ );  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 50.03, 109.06, 113.05, 115.29, 122.42, 125.13, 125.73, 125.77, 125.80, 125.84, 128.78, 129.26, 138.28, 144.78, 148.47, 196.02; Anal. Calcd. for  $\text{C}_{15}\text{H}_{13}\text{F}_3\text{N}_2\text{O}_3\text{S}$ : C 50.28; H 3.66; N 7.82 %. Found: C 50.28; H 3.62; N 7.80 %.

### General procedure for the synthesis of imidazoles **10** and **13**.

An amount of 2 mmol of compounds **3**, **6** was dissolved in the solution of glacial acetic acid (5 mL) and HCl (1 mL), and KSCN (0.78 g, 8 mmol) was added. The reaction mixture was heated at reflux for 4 h, then it was cooled down, diluted with water, and the precipitate was filtered off and washed with water and n-hexane.

**3-(4-(4-bromophenyl)-2-thioxo-2,3-dihydro-1H-imidazol-1-yl)benzenesulfonamide (10)**. Yellowish solid, yield 0.59 g (72%); m.p. 280–281 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3258, 2729, 1485;  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 7.52 (s, 2H,  $\text{NH}_2$ ), 7.60–7.79 (m, 5H,  $H_{\text{ar}}$ ), 7.84–7.98 (m, 2H,  $H_{\text{ar}}$ ), 8.02 (s, 1H,  $H_{\text{ar}}$ ), 8.18 (s, 1H, CH), 13.12 (s, 1H, SH);  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 116.38, 120.95, 122.98, 125.01, 126.26, 126.86, 127.65, 129.25, 129.68, 131.81, 137.79, 144.76, 163.17; Anal. Calcd. For  $\text{C}_{15}\text{H}_{12}\text{BrN}_3\text{O}_2\text{S}_2$ : C 43.91; H 2.95; N 10.24 %. Found: C 43.99; H 2.91; N 10.21 %.

**3-(2-thioxo-4-(4-(trifluoromethyl)phenyl)-2,3-dihydro-1H-imidazol-1-yl)benzenesulfonamide (13)**. Light brown solid, yield 0.51 g (75%); m.p. 226–227 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3141, 2737, 1489;  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 7.53 (s, 2H,  $\text{NH}_2$ ), 7.72–8.05 (m, 7H,  $H_{\text{ar}}$ ), 8.13–8.24 (m, 2H, CH,  $H_{\text{ar}}$ ), 13.26 (s, 1H, SH);  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 117.77, 123.06, 124.75, 125.14, 125.91, 125.95, 127.24, 127.67, 127.99, 129.32, 129.71, 131.53, 137.70, 144.80, 163.62; Anal. Calcd. For  $\text{C}_{16}\text{H}_{12}\text{F}_3\text{N}_3\text{O}_2\text{S}_2$ : C 48.12; H 3.03; N 10.52 %. Found: C 48.10; H 2.99; N 10.47 %.

### General procedure for the synthesis of S-alkylated compounds **16–22a-c**.

Imidazole **9–15** (1.0 mmol) was dissolved in DMF (3 mL). Triethylamine (0.5 mL) and corresponding alkyl halide (1.5 mmol) were added dropwise, and the reaction mixture was stirred at room temperature for 2–3 h. Then, the reaction mixture was diluted with 20 mL of water. The precipitate was filtered off, washed with water and diethyl ether, dried, and recrystallized from propan-2-ol.

**3-(2-(methylthio)-4-phenyl-1H-imidazol-1-yl)benzenesulfonamide (16a).** White solid, yield 0.29 g (84%); m.p. 148–149 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3327, 1483;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 2.63 (s, 3H,  $\text{CH}_3$ ), 7.25 (t, 1H,  $J = 7.4$  Hz,  $\text{H}_{\text{ar}}$ ), 7.23 (t, 2H,  $J = 7.6$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.75–7.98 (m, 6H,  $\text{H}_{\text{ar}}$ ), 8.08 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 14.83, 27.56, 115.34, 115.56, 118.69, 122.32, 125.44, 126.21, 126.29, 128.63, 130.00, 130.03, 130.47, 136.94, 140.57, 141.88, 145.36; Anal. Calcd. For  $\text{C}_{16}\text{H}_{15}\text{N}_3\text{O}_3\text{S}_2$ : C 55.63; H 4.38; N 12.16; %. Found: C 55.57; H 4.35; N 12.13 %.

**3-(2-(ethylthio)-4-phenyl-1H-imidazol-1-yl)benzenesulfonamide (16b).** Light yellow solid, yield 0.27 g (75%); m.p. 160–161 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3324, 1482;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 1.30 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 3.14 (q, 2H,  $J = 7.3$  Hz,  $\text{CH}_2$ ), 7.23 (t, 2H,  $J = 8.7$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.70–8.00 (m, 6H,  $\text{H}_{\text{ar}}$ ), 8.08 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 14.84, 27.59, 118.84, 122.34, 124.38, 125.40, 126.87, 128.59, 128.64, 130.46, 133.43, 136.99, 141.46, 141.79, 145.35; Anal. Calcd. For  $\text{C}_{17}\text{H}_{17}\text{N}_3\text{O}_2\text{S}_2$ : C 56.80; H 4.77; N 11.69; %. Found: C 56.79; H 4.75; N 11.64 %.

**3-(4-phenyl-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide (16c).** White solid, yield 0.30 g (81%); m.p. 130–131 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3332, 1483;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 0.92 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 1.67 (extette, 2H,  $J = 7.3$  Hz,  $\text{CH}_2$ ), 3.12 (t, 2H,  $J = 7.1$  Hz,  $\text{SCH}_2$ ), 7.25 (t, 1H,  $J = 7.4$  Hz,  $\text{H}_{\text{ar}}$ ), 7.40 (t, 2H,  $J = 7.6$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.73–7.87 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.94 (br. s, 2H,  $\text{H}_{\text{ar}}$ ), 8.08 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 13.04, 22.41, 35.18, 118.85, 122.36, 124.36, 125.40, 126.86, 128.58, 128.65, 130.45, 133.43, 137.00, 141.40, 141.92, 145.36; Anal. Calcd. For  $\text{C}_{18}\text{H}_{19}\text{N}_3\text{O}_2\text{S}_2$ : C 57.89; H 5.13; N 11.25; %. Found: C 57.88; H 5.09; N 11.29 %.

**3-(4-(4-bromophenyl)-2-(methylthio)-1H-imidazol-1-yl)benzenesulfonamide (17a).** White solid, yield 0.36 g (85%); m.p. 168–169 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3351, 1480;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 2.63 (s, 3H,  $\text{CH}_3$ ), 7.54–7.62 (m, 4H,  $\text{NH}_2$ ,  $\text{H}_{\text{ar}}$ ), 7.76–7.84 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.94 (br. s, 2H,  $\text{H}_{\text{ar}}$ ), 8.16 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 15.44, 119.39, 119.60, 122.11, 125.52, 126.32, 128.42, 130.58, 131.50, 132.70, 136.77, 140.23, 143.35, 145.45; Anal. Calcd. For  $\text{C}_{16}\text{H}_{14}\text{BrN}_3\text{O}_2\text{S}_2$ : C 45.29; H 3.33; N 9.90; %. Found: C 45.25; H 3.29; N 9.90 %.

**3-(4-(4-bromophenyl)-2-(ethylthio)-1H-imidazol-1-yl)benzenesulfonamide (17b).** White solid, yield 0.38 g (80%); m.p. 142–143 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3350, 1479;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 1.30 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 3.15 (q, 2H,  $J = 7.3$  Hz,  $\text{CH}_2$ ), 7.53–7.61 (m, 4H,  $\text{NH}_2$ ,  $\text{H}_{\text{ar}}$ ), 7.73–7.83 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.90–7.96 (m, 2H,  $\text{H}_{\text{ar}}$ ), 8.16 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 14.82, 27.50, 119.42, 119.63, 122.32, 125.50, 126.34, 128.64, 130.49, 131.52, 132.72, 136.86, 140.31, 142.20, 145.35; Anal. Calcd. For  $\text{C}_{17}\text{H}_{16}\text{BrN}_3\text{O}_2\text{S}_2$ : C 46.58; H 3.68; N 9.59; %. Found: C 46.61; H 3.69; N 9.55 %.

**3-(4-(4-bromophenyl)-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide (17c).** White solid, yield 0.39 g (75%); m.p. 163–164 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3352, 1479;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 0.92 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 1.66 (extette, 2H,  $J = 7.2$  Hz,  $\text{CH}_2$ ), 3.13 (t, 2H,  $J = 7.1$  Hz,  $\text{SCH}_2$ ), 7.52–7.63 (m, 4H,  $\text{NH}_2$ ,  $\text{H}_{\text{ar}}$ ), 7.73–7.84 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.89–7.96 (m, 2H,  $\text{H}_{\text{ar}}$ ), 8.15 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 13.04, 22.37, 35.10, 119.42, 119.63, 122.35, 125.50, 126.32, 128.65, 130.48, 131.52, 132.71, 136.87, 140.25, 142.34, 145.38; Anal. Calcd. For  $\text{C}_{18}\text{H}_{18}\text{BrN}_3\text{O}_2\text{S}_2$ : C 47.79; H 4.01; N 9.29; %. Found: C 47.76; H 3.98; N 9.25 %.

**3-(4-(4-chlorophenyl)-2-(methylthio)-1H-imidazol-1-yl)benzenesulfonamide (18a).** Brownish solid, yield 0.28 g (74%); m.p. 171–172 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3347, 1482;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 2.63 (s, 3H,  $\text{CH}_3$ ), 7.45 (d, 2H,  $J = 8.2$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.74–7.90 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.95 (br. s, 2H,  $\text{H}_{\text{ar}}$ ), 8.14 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 15.45, 119.33, 122.12, 125.52, 126.00, 128.42, 128.61, 130.59, 131.12, 132.35, 136.79, 140.22, 143.33, 145.46; Anal. Calcd. For  $\text{C}_{16}\text{H}_{14}\text{ClN}_3\text{O}_2\text{S}_2$ : C 50.59; H 3.71; N 11.06; %. Found: C 50.58; H 3.68; N 11.01 %.

**3-(4-(4-chlorophenyl)-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide (18c).** Light brown solid, yield 0.35 g (85%); m.p. 166–167 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3348, 1481;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  (ppm): 0.93 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 1.67 (sextet, 2H,

$J = 7.3$  Hz,  $\text{CH}_2$ ), 3.13 (t, 2H,  $J = 7.1$  Hz,  $\text{SCH}_2$ ), 7.46 (d, 2H,  $J = 8.3$  Hz,  $\text{H}_{\text{ar}}$ ), 7.58 (br. s, 2H,  $\text{NH}_2$ ), 7.74–7.99 (m, 6H,  $\text{H}_{\text{ar}}$ ), 8.15 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 13.04, 22.38, 35.10, 119.37, 122.35, 125.50, 126.00, 128.62, 128.65, 130.48, 131.13, 132.36, 136.88, 140.23, 142.31, 145.38; Anal. Calcd. For  $\text{C}_{18}\text{H}_{18}\text{ClN}_3\text{O}_2\text{S}_2$ : C 53.00; H 4.45; N 10.30; %. Found: C 52.99; H 4.39; N 10.30 %.

**3-(4-(4-fluorophenyl)-2-(methylthio)-1H-imidazol-1-yl)benzenesulfonamide (19a).** Yellowish solid, yield 0.29 g (81%); m.p. 162–163 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3341, 1484;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 2.63 (s, 3H,  $\text{CH}_3$ ), 7.23 (t, 2H,  $J = 8.7$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.75–7.98 (m, 6H,  $\text{H}_{\text{ar}}$ ), 8.08 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 15.50, 115.33, 115.55, 118.64, 122.10, 125.45, 126.20, 126.28, 128.40, 130.00, 130.03, 130.57, 136.86, 140.51, 143.04, 145.45; Anal. Calcd. For  $\text{C}_{16}\text{H}_{14}\text{FN}_3\text{O}_2\text{S}_2$ : C 52.88; H 3.88; N 11.56; %. Found: C 52.85; H 3.89; N 11.51 %.

**3-(2-(ethylthio)-4-(4-fluorophenyl)-1H-imidazol-1-yl)benzenesulfonamide (19b).** Yellowish solid, yield 0.31 g (82%); m.p. 160–161 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3350, 1483;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 1.30 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 3.14 (q, 2H,  $J = 7.3$  Hz,  $\text{CH}_2$ ), 7.24 (t, 2H,  $J = 8.7$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.69–7.99 (m, 6H,  $\text{H}_{\text{ar}}$ ), 8.09 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 14.84, 27.59, 115.36, 115.57, 118.72, 122.33, 125.46, 126.22, 126.30, 128.64, 129.97, 130.48, 136.92, 140.51, 141.88, 145.37; Anal. Calcd. For  $\text{C}_{17}\text{H}_{16}\text{FN}_3\text{O}_2\text{S}_2$ : C 54.10; H 4.27; N 11.13; %. Found: C 54.05; H 4.23; N 11.09 %.

**3-(4-(4-fluorophenyl)-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide (19c).** White solid, yield 0.30 g (77%); m.p. 142–143 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3350, 1484;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 0.92 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 1.66 (sextet, 2H,  $J = 7.2$  Hz,  $\text{CH}_2$ ), 3.12 (t, 2H,  $J = 7.1$  Hz,  $\text{SCH}_2$ ), 7.23 (t, 2H,  $J = 8.6$  Hz,  $\text{H}_{\text{ar}}$ ), 7.57 (br. s, 2H,  $\text{NH}_2$ ), 7.72–7.96 (m, 6H,  $\text{H}_{\text{ar}}$ ), 8.07 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 13.04, 22.40, 35.15, 115.35, 115.56, 118.70, 122.33, 125.43, 126.19, 126.27, 128.64, 130.00, 130.03, 130.47, 136.95, 140.51, 142.01, 145.36; Anal. Calcd. For  $\text{C}_{18}\text{H}_{18}\text{FN}_3\text{O}_2\text{S}_2$ : C 55.23; H 4.63; N 10.73; %. Found: C 55.19; H 4.57; N 10.76 %.

**3-(2-(methylthio)-4-(4-(trifluoromethyl)phenyl)-1H-imidazol-1-yl)benzenesulfonamide (20a).** Yellow solid, yield 0.34 g (83%); m.p. 206–207 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3341, 1484;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 2.65 (s, 3H,  $\text{CH}_3$ ), 7.58 (br. s, 2H,  $\text{NH}_2$ ), 7.71–7.84 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.91–8.12 (m, 4H,  $\text{H}_{\text{ar}}$ ), 8.30 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 15.41, 120.65, 122.18, 123.11, 124.70, 125.53, 125.57, 125.61, 125.66, 125.81, 126.72, 127.03, 128.48, 130.62, 136.68, 137.39, 139.87, 143.88, 145.50; Anal. Calcd. For  $\text{C}_{17}\text{H}_{14}\text{F}_3\text{N}_3\text{O}_2\text{S}_2$ : C 49.39; H 3.41; N 10.16; %. Found: C 49.31; H 3.43; N 10.11 %.

**3-(2-(ethylthio)-4-(4-(trifluoromethyl)phenyl)-1H-imidazol-1-yl)benzenesulfonamide (20b).** White solid, yield 0.33 g (77%); m.p. 154–155 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3334, 1483;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 1.32 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 3.17 (q, 2H,  $J = 7.3$  Hz,  $\text{CH}_2$ ), 7.58 (br. s, 2H,  $\text{NH}_2$ ), 7.70–7.85 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.95 (br. s, 2H,  $\text{H}_{\text{ar}}$ ), 8.05 (d, 2H,  $J = 8.1$  Hz,  $\text{H}_{\text{ar}}$ ), 8.30 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 14.80, 27.49, 120.65, 122.39, 123.10, 124.73, 125.55, 125.59, 125.64, 125.81, 126.75, 127.07, 128.69, 130.53, 136.77, 137.42, 139.95, 142.74, 145.42; Anal. Calcd. For  $\text{C}_{18}\text{H}_{16}\text{F}_3\text{N}_3\text{O}_2\text{S}_2$ : C 50.58; H 3.77; N 9.83; %. Found: C 50.55; H 3.72; N 9.79 %.

**3-(2-(propylthio)-4-(4-(trifluoromethyl)phenyl)-1H-imidazol-1-yl)benzenesulfonamide (20c).** Yellowish solid, yield 0.38 g (86%); m.p. 133–134 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3352, 1486;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 0.93 (t, 3H,  $J = 7.3$  Hz,  $\text{CH}_3$ ), 1.68 (sextet, 2H,  $J = 7.3$  Hz,  $\text{CH}_2$ ), 3.15 (t, 2H,  $J = 7.1$  Hz,  $\text{SCH}_2$ ), 7.58 (br. s, 2H,  $\text{NH}_2$ ), 7.70–7.84 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.95 (br. s, 2H,  $\text{H}_{\text{ar}}$ ), 8.05 (d, 2H,  $J = 8.1$  Hz,  $\text{H}_{\text{ar}}$ ), 8.29 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm): 13.04, 22.36, 35.08, 120.66, 122.41, 123.10, 124.71, 125.55, 125.59, 125.64, 125.80, 126.75, 127.06, 128.71, 130.52, 136.78, 137.41, 139.89, 142.87, 145.43; Anal. Calcd. For  $\text{C}_{19}\text{H}_{18}\text{F}_3\text{N}_3\text{O}_2\text{S}_2$ : C 51.69; H 4.11; N 9.52; %. Found: C 51.63; H 4.09; N 9.50 %.

**3-(4-(4-cyanophenyl)-2-(methylthio)-1H-imidazol-1-yl)benzenesulfonamide (21a).** Light yellow solid, yield 0.26 g (70%); m.p. 192–193 °C; IR (KBr) ( $\nu$ ,  $\text{cm}^{-1}$ ): 3336, 1484;  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$  (ppm): 2.65 (s, 3H,  $\text{CH}_3$ ), 7.58 (br. s, 2H,  $\text{NH}_2$ ), 7.77–7.89 (m, 4H,  $\text{H}_{\text{ar}}$ ), 7.93–8.05 (m, 4H,  $\text{H}_{\text{ar}}$ ), 8.34 (s, 1H, CH);  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-d}_6$ ) ( $\delta$ , ppm):

15.36, 108.76, 119.15, 121.37, 122.19, 124.77, 125.74, 128.50, 130.65, 132.70, 136.59, 137.93, 139.63, 144.22, 145.51; Anal. Calcd. For  $C_{17}H_{14}N_4O_2S_2$ : C 55.12; H 3.81; N 15.12; %. Found: C 55.07; H 3.78; N 15.13 %.

**3-(4-(4-cyanophenyl)-2-(ethylthio)-1H-imidazol-1-yl)benzenesulfonamide (21b).** Light yellow solid, yield 0.29 g (76%); m.p. 204–205 °C; IR (KBr) ( $\nu$ ,  $cm^{-1}$ ): 3350, 1483;  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 1.32 (t, 3H,  $J = 7.3$  Hz,  $CH_3$ ), 3.17 (q, 2H,  $J = 7.3$  Hz,  $CH_2$ ), 7.58 (br. s, 2H,  $NH_2$ ), 7.74–8.09 (m, 8H,  $H_{ar}$ ), 8.34 (s, 1H, CH);  $^{13}C$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 14.80, 27.44, 108.78, 119.15, 121.37, 122.38, 124.78, 125.72, 128.71, 130.57, 132.72, 136.67, 137.95, 139.69, 143.09, 145.44; Anal. Calcd. For  $C_{18}H_{16}N_4O_2S_2$ : C 56.23; H 4.19; N 14.57; %. Found: C 56.17; H 4.15; N 14.59 %.

**3-(4-(4-cyanophenyl)-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide (21c).** White solid, yield 0.27 g (68%); m.p. 210–211 °C; IR (KBr) ( $\nu$ ,  $cm^{-1}$ ): 3351, 1483;  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 0.93 (t, 3H,  $J = 7.3$  Hz,  $CH_3$ ), 1.68 (sextet, 2H,  $J = 7.2$  Hz,  $CH_2$ ), 3.15 (t, 2H,  $J = 7.1$  Hz,  $SCH_2$ ), 7.58 (br. s, 2H,  $NH_2$ ), 7.75–8.07 (m, 8H,  $H_{ar}$ ), 8.33 (s, 1H, CH);  $^{13}C$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 13.05, 22.37, 35.01, 108.78, 119.15, 121.37, 122.40, 124.77, 125.73, 128.71, 130.56, 132.72, 136.68, 137.94, 139.64, 143.23, 145.44; Anal. Calcd. For  $C_{19}H_{18}N_4O_2S_2$ : C 57.27; H 4.55; N 14.06; %. Found: C 57.21; H 4.50; N 14.08 %.

**3-(2-(methylthio)-4-(4-nitrophenyl)-1H-imidazol-1-yl)benzenesulfonamide (22a).** Yellow solid, yield 0.31 g (79%); m.p. 204–205 °C; IR (KBr) ( $\nu$ ,  $cm^{-1}$ ): 3264, 1489;  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 2.66 (s, 3H,  $CH_3$ ), 7.58 (br. s, 2H,  $NH_2$ ), 7.78–7.84 (m, 2H,  $H_{ar}$ ), 7.97 (br. s, 2H,  $H_{ar}$ ), 8.09 (d, 2H,  $J = 8.4$  Hz,  $H_{ar}$ ), 8.28 (d, 2H,  $J = 8.4$  Hz,  $H_{ar}$ ), 8.42 (s, 1H, CH);  $^{13}C$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 15.34, 122.12, 122.19, 124.23, 124.85, 125.81, 128.51, 130.67, 136.51, 139.30, 140.00, 144.63, 145.52, 145.71; Anal. Calcd. For  $C_{16}H_{14}N_4O_2S_2$ : C 49.22; H 3.61; N 14.35; %. Found: C 49.24; H 3.57; N 14.37 %.

**3-(2-(ethylthio)-4-(4-nitrophenyl)-1H-imidazol-1-yl)benzenesulfonamide (22b).** Yellow solid, yield 0.33 g (81%); m.p. 188–189 °C; IR (KBr) ( $\nu$ ,  $cm^{-1}$ ): 3312, 1484;  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 1.33 (t, 3H,  $J = 7.3$  Hz,  $CH_3$ ), 3.19 (q, 2H,  $J = 7.3$  Hz,  $CH_2$ ), 7.59 (br. s, 2H,  $NH_2$ ), 7.74–7.86 (m, 2H,  $H_{ar}$ ), 7.96 (br. s, 2H,  $H_{ar}$ ), 8.09 (d, 2H,  $J = 8.5$  Hz,  $H_{ar}$ ), 8.27 (d, 2H,  $J = 8.5$  Hz,  $H_{ar}$ ), 8.42 (s, 1H, CH);  $^{13}C$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 14.78, 27.43, 122.08, 122.38, 124.23, 124.87, 125.79, 128.70, 130.58, 136.60, 139.37, 140.01, 143.50, 145.46, 145.73; Anal. Calcd. For  $C_{17}H_{16}N_4O_2S_2$ : C 50.48; H 3.99; N 13.85; %. Found: C 50.49; H 3.99; N 13.79 %.

**3-(4-(4-nitrophenyl)-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide (22c).** Yellow solid, yield 0.34 g (81%); m.p. 174–175 °C; IR (KBr) ( $\nu$ ,  $cm^{-1}$ ): 3314, 1484;  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 0.94 (t, 3H,  $J = 7.3$  Hz,  $CH_3$ ), 1.69 (sextet, 2H,  $J = 7.3$  Hz,  $CH_2$ ), 3.17 (t, 2H,  $J = 7.1$  Hz,  $SCH_2$ ), 7.58 (br. s, 2H,  $NH_2$ ), 7.76–7.85 (m, 2H,  $H_{ar}$ ), 7.96 (br. s, 2H,  $H_{ar}$ ), 8.08 (d, 2H,  $J = 8.5$  Hz,  $H_{ar}$ ), 8.28 (d, 2H,  $J = 8.5$  Hz,  $H_{ar}$ ), 8.41 (s, 1H, CH);  $^{13}C$  NMR (101 MHz, DMSO- $d_6$ ) ( $\delta$ , ppm): 13.06, 22.33, 35.00, 122.12, 122.40, 124.24, 124.86, 125.79, 128.73, 130.58, 136.61, 139.31, 140.00, 143.62, 145.45, 145.73; Anal. Calcd. For  $C_{18}H_{18}N_4O_2S_2$ : C 51.66; H 4.34; N 13.39; %. Found: C 51.62; H 4.29; N 13.37 %.

### 2.3. Minimal Inhibitory Concentration Determination

#### 2.3.1. Preparation of Assay Microplates

The minimal inhibitory concentrations (MICs) of compounds **2–22a–c**, as well as of clinically approved antibiotics (rifampin, isoniazid, amikacin, levofloxacin, and meropenem) were determined by microplate broth dilution method as described by Clinical Laboratory Standards Institute document M07-A8. The antimicrobials were selected to represent major antimicrobials used in clinical settings to treat MDR infections, as well as infections caused by rapidly growing *Mycobacterium* spp. The MICs for the compounds and comparator antibiotics were determined against the libraries of Gram-positive and Gram-negative pathogens, multidrug-resistant fungi, and mycobacteria.

Compounds and antibiotics that were used as a control were dissolved in molecular biology grade dimethyl sulfoxide (DMSO) to achieve a final concentration of 25–30 mg/mL. Compound dilutions were achieved in 1.5 mL polypropylene 96-well microplates to gener-



ate 2× of concentrations of each drug (0.5–64 µg/mL). The 2× concentrates were then transferred to flat bottom plates and used for inoculation or stored in argon purged sealed bags at −80 °C.

### 2.3.2. Antibacterial Activity Characterization Using Gram-Positive and Gram-Negative Pathogens

A microbial inoculum was prepared using the direct colony suspension method and densitometric analysis. The inoculum suspension of each test organism was prepared in 5 mL of sterile deionized water until densitometer reached 0.5 MFA and further diluted in sterile CAMBH media to achieve final concentrations of approximately  $5 \times 10^5$  CFU/mL in each well after dispensing in microplates. The inoculum was transferred to the assay plates to achieve 1× assay concentration. A 10 µL of inoculum was plated on Sheep Blood agar plates to validate the purity and inoculum size. Inoculated microdilution plates were incubated at 35 °C for 16 to 20 h in an ambient-air incubator.

### 2.3.3. Antifungal Activity Characterization

The MIC of compounds 2–22a–c, as well as clinically approved antifungal drugs was determined by CLSI recommendations that were described in document M27-A3 [36,37]. Multidrug-resistant *Candida* spp. strains were sub-cultured on Sabouraud-Dextrose agar for 24 h at 35 °C. Drug-resistant *Aspergillus fumigatus* was cultured on Inhibitory mold agar slants for 5 days at 35 °C. The colonies of *Candida* isolates were suspended in sterile saline to reach approximately  $5 \times 10^6$  CFU/mL. The conidia of *A. fumigatus* were collected by flooding the slants with saline containing 0.5% of Tween 80 and passing conidia through 75 µm cell strainer. The inoculums were quantified by using haematocytometer and then, the fungal suspension was diluted in RPMI/MOPS broth to reach  $5 \times 10^5$  CFU/mL. The inoculum was then dispensed in assay microplates, and inoculated microdilution plates were incubated at 35 °C for 24 h in an ambient-air incubator within 15 min of the addition of the inoculum.

### 2.3.4. Antimycobacterial Activity Determination

Before the experiments, multidrug-resistant *M. abscessus* complex strains were cultured on Middlebrook 7H9 agar containing ODAC supplement for four days at 37 °C. *M. bovis* BCG and avirulent *M. tuberculosis* H37Ra strains were grown on Lowenstein–Jensen (LJ) media for 3 weeks.

The colonies of *M. abscessus* were scraped and suspended in tube with sterile saline to achieve approximately  $5 \times 10^6$  CFU/mL. *M. bovis* BCG and *M. tuberculosis* H37Ra were scraped from the LJ media and transferred to the tube containing 4 mL of Middlebrook 7H9 broth and 3 borosilicate glass beads. The tube was vortexed on maximum speed for 2 min, and then, the bacterial suspension was adjusted to  $5 \times 10^6$  CFU/mL. Prior to inoculation of the plates, the bacterial suspension was diluted 1:10 in Middlebrook 7H9 broth containing 20 µg/mL of resazurin, and microplates were inoculated by using multichannel pipette.

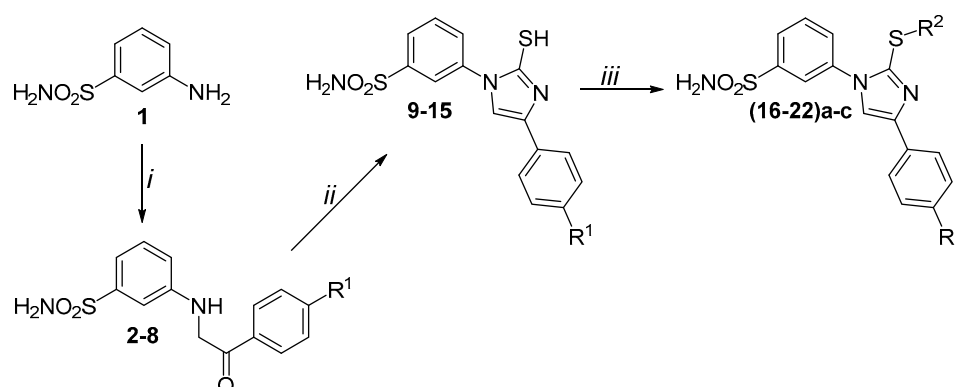
The plates were incubated at 37 °C in humidified incubator for 5 days (for *M. abscessus* complex) or two weeks (for *M. bovis* BCG and *M. tuberculosis* H37Ra) and the minimal inhibitory concentration was determined by visual evaluation.

## 3. Results and Discussion

### 3.1. Chemistry

Most of the compounds 2–15 (Scheme 1) were resynthesized according to our previous study [35] and were further investigated during this study. All the spectral data and reaction conditions can be found in previously mentioned research [35]. Moreover, to explore further on benzenesulfonamide-bearing 1H-imidazolethiol moieties, new compounds 3, 6, 10, and 13 were newly synthesized for this study (Scheme 1). 3-Aminobenzenesulfonamide (1) was treated with various α-halogenketones in water/1,4-dioxane solution to afford compound 3 and 6. These intermediate compounds were later cyclized with potassium thiocyanate

in glacial acetic acid and in a presence of HCl as a catalyst into 1H-imidazole derivatives **10** and **13**. The structures of compounds **3**, **6**, **10**, and **13** have also been confirmed by the data of FT-IR,  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopy, as well as elemental analysis data. For instance, in a  $^1\text{H}$  NMR spectrum for **10**, the singlets assigned to the protons in the CH group at 8.18 ppm and in the SH group at 13.12 ppm have proven the presence of 1H-imidazolethiol moiety in the molecule. One of the best-known properties of thioamides is the tautomerism [38]: thioamides can exist in their thione/thiol forms. However, the  $^{13}\text{C}$  NMR spectral data showed that in DMSO- $d_6$  solvent, thiol tautomeric form is predominant for both compounds **10** and **13**. The carbon attributed to the C-SH group resonated at 163.17 and 163.62 ppm, respectively.



**2:**  $\text{R}^1 = \text{H}$ ; **3:**  $\text{R}^1 = \text{Br}$ ; **4:**  $\text{R}^1 = \text{Cl}$ ; **5:**  $\text{R}^1 = \text{F}$ ; **6:**  $\text{R}^1 = \text{CF}_3$ ; **7:**  $\text{R}^1 = \text{CN}$ ; **8:**  $\text{R}^1 = \text{NO}_2$ ; **9:**  $\text{R}^1 = \text{H}$ ;  
**10:**  $\text{R}^1 = \text{Br}$ ; **11:**  $\text{R}^1 = \text{Cl}$ ; **12:**  $\text{R}^1 = \text{F}$ ; **13:**  $\text{R}^1 = \text{CF}_3$ ; **14:**  $\text{R}^1 = \text{CN}$ ; **15:**  $\text{R}^1 = \text{NO}_2$ ; **16:**  $\text{R}^1 = \text{H}$ ;  
**17:**  $\text{R}^1 = \text{Br}$ ; **18:**  $\text{R}^1 = \text{Cl}$ ; **19:**  $\text{R}^1 = \text{F}$ ; **20:**  $\text{R}^1 = \text{CF}_3$ ; **21:**  $\text{R}^1 = \text{CN}$ ; **22:**  $\text{R}^1 = \text{NO}_2$ ;  
**a:**  $\text{R}^2 = \text{CH}_3$ ; **b:**  $\text{R}^2 = \text{CH}_2\text{CH}_3$ ; **c:**  $\text{R}^2 = \text{CH}_2\text{CH}_2\text{CH}_3$ ;

Reaction conditions: *i*) corresponding  $\alpha$ -halogenketone, water, 1,4-dioxane, reflux, 2h; *ii*) KSCN, HCl, glacial acetic acid, reflux, 4h; *iii*) corresponding alkyl halide, TEA, DMF, r.t., 2-3h;

**Scheme 1.** Synthesis of compounds (2-22)a-c.

The main goal of this study was to further investigate 1H-imidazolethiol derivatives with various alkyl substituents. For this purpose, S-alkylation reactions with bromomethane, ethyl iodide, and n-propyl iodide in dimethyl formamide were carried out to obtain compounds **16-22a-c**. Triethylamine was used as a base catalyst to increase the reaction rate. For example, in a  $^1\text{H}$  NMR spectrum for **16a**, the singlet assigned to the protons in the  $\text{CH}_3$  group at 2.63 ppm have proved the presence of methyl moiety in the molecule, while a triplet at 0.92 ppm, a sextet at 1.67 ppm, and a triplet at 3.12 ppm assigned to the protons in the  $\text{CH}_3$ ,  $\text{CH}_2$ , and  $\text{CH}_3$ , respectively, proved the presence of a propyl group in compound **16c**. Elemental analysis data of compounds **16-22a-c** confirmed that all the molecules did not form hydroiodide or hydrobromide salts.

### 3.2. Benzenesulfonamide Derivatives 2-22a-c Demonstrated Structure-Depended Antimicrobial Activity against Multidrug-Resistant Non-Tuberculous Mycobacteria

Novel benzenesulfonamide derivatives bearing substituted imidazoles demonstrated structure-depended antimicrobial activity against *Mycobacterium abscessus* complex strains (Table 1). Notably, compounds **2-22a-c** showed little activity against multidrug-resistant Gram-positive and Gram-negative bacterial strains or drug-resistant fungi, suggesting the mycobacteria-directed activity (Tables S1 and S2).



**Table 1.** The in vitro antimicrobial activity of compounds 2–22a–c against mycobacteria. The minimal inhibitory concentration (MIC) values are provided as an average value obtained from three experimental replicas.

Compound	Minimal Inhibitory Concentration (µg/mL)						
	<i>M. abscessus</i> MA1884	<i>M. abscessus</i> MA1753	<i>M. abscessus</i> MA1836	<i>M. abscessus</i> MA1704	<i>M. abscessus</i> MA0040	<i>M. bovis</i> BCG	<i>M. tuberculosis</i> H37Ra
2	>64	>64	>64	>64	>64	>64	>64
3	>64	>64	>64	>64	>64	>64	>64
4	>64	>64	>64	>64	>64	>64	>64
5	>64	>64	>64	>64	>64	>64	>64
6	64	64	>4	64	64	>64	>64
7	>64	>64	>64	>64	>64	>64	>64
8	>64	>64	>64	>64	>64	>64	>64
9	64	64	64	>64	>64	32	32
10	>64	>64	>64	>64	>64	>64	>64
11	>64	>64	>64	>64	>64	>64	>64
12	64	64	32	64	>64	>64	>64
13	1	4	4	4	4	1	0.5
14	>64	>64	>64	>64	>64	>64	>64
15	>64	>64	>64	>64	>64	>64	>64
16a	64	64	>64	>64	>64	>64	>64
16b	>64	>64	>64	>64	>64	>64	>64
16c	>64	>64	>64	>64	>64	>64	>64
17a	>64	>64	>64	>64	>64	>64	>64
17b	>64	>64	>64	>64	>64	>64	>64
17c	>64	>64	>64	>64	>64	>64	>64
18a	>64	>64	>64	>64	>64	>64	>64
18b	64	64	>64	>64	32	>64	>64
18c	64	64	32	16	32	32	16
19a	8	8	4	4	8	4	4
19b	>64	>64	>64	>64	>64	>64	>64
19c	>64	>64	>64	>64	>64	>64	>64
20a	>64	>64	>64	>64	>64	>64	>64
20b	>64	>64	>64	>64	>64	>64	>64
20c	>64	>64	>64	>64	>64	>64	>64
21a	>64	>64	>64	>64	>64	>64	>64
21b	>64	>64	>64	>64	>64	>64	>64
21c	>64	>64	>64	>64	>64	>64	>64
22a	>64	>64	>64	>64	>64	>64	>64
22b	>64	>64	>64	>64	>64	>64	>64
22c	64	64	>64	>64	64	>64	>64
Rifampin	64	>64	32	16	32	≤0.5	0.5
Isoniazid	8	4	16	32	32	0.5	≤0.5
Amikacin	32	32	16	16	32	≤0.5	≤0.5
Levofloxacin	16	8	16	32	32	1	1
Meropenem	8	8	32	32	64	8	4

Compounds 2–5, bearing 4-H or halogen substitutions demonstrated no antimicrobial activity against *M. abscessus* complex strains, as well as *M. bovis* BCG or *M. tuberculosis* H37Ra (MIC > 64 µg/mL). The addition of the 4-CF<sub>3</sub> substitution on the benzenesulfonamide core in compound 6 resulted in weak antimicrobial activity against *M. abscessus* complex strains (MIC 64 µg/mL) except for *M. abscessus* MA1836. Moreover, compound 7 showed no activity against *M. bovis* BCG or *M. tuberculosis* H37Ra strains (MIC > 64 µg/mL). Furthermore, the incorporation of 4-CN (7), or 4-NO<sub>2</sub> (8), in the benzenesulfonamide nucleus diminished the antimicrobial activity against *M. abscessus* complex, as well as *M. bovis* BCG or *M. tuberculosis* H37Ra (Table 1).

The incorporation of imidazole-2-thiol moiety in compound 9 resulted in weak antimicrobial activity against *M. abscessus* complex strains (MIC 64 µg/mL) with exception

of *M. abscessus* MA1704 and MA0040 (MIC > 64 µg/mL). The incorporation of imidazole-2-thiol moiety (compound **9**) resulted in extended antimicrobial activity against rapidly growing *M. abscessus* strains, non-tuberculous mycobacteria (*M. bovis* BCG), as well as *M. tuberculosis* H37Ra (MIC 32 µg/mL, respectively). Interestingly, 4-Br, 4-Cl substitutions in imidazole-2-thiol derivatives (**10,11**) resulted in loss of antimicrobial activity against mycobacteria, while 4-F substitution (**12**) resulted in antimicrobial activity against *M. abscessus* complex (MIC 32–64 µg/mL) and loss of activity against *M. bovis* BCG and *M. tuberculosis* H37Ra (MIC > 64 µg/mL). The further addition of 4-CF<sub>3</sub> substitution resulted in compound **13** with strong antimicrobial activity against tested mycobacterial strains (MIC 0.5–4 µg/mL). The antimicrobial activity of compound **13** against *M. abscessus* complex was greater than rifampicin (MIC 32–64 µg/mL), isoniazid (MIC 4–32 µg/mL), amikacin (MIC 16–32 µg/mL), levofloxacin (MIC 8–32 µg/mL), and meropenem (MIC 8–64 µg/mL) (Table 1).

The incorporation of an aryl group often results in increased lipophilicity of the compounds. Therefore, we further postulated that the incorporation of various length aryl substitutions if benzenesulfonamide derivatives could enhance the mycobacteria-directed antimicrobial activity. Compound **16a** bearing methyl group demonstrated weak antimicrobial activity against *M. abscessus* complex strains MA1884 and MA1753 (MIC 64 µg/mL). The elongation of the aryl chain by adding ethyl and propyl groups (**16b** and **16c**) diminished the antimicrobial activity. On the other hand, compounds **18a–c** containing the 4-Cl substitution demonstrated that the length of the aryl chain is mediating the antimicrobial activity. Compound **18a** bearing the methyl substitution showed no antimicrobial activity while compound **18b** containing the ethyl group showed antimicrobial activity against *M. abscessus* complex (MIC 32–64 µg/mL), but not *M. bovis* BCG or *M. tuberculosis* H37Ra (MIC > 64 µg/mL). Notably, the incorporation of propyl substitution (**18c**) resulted in enhanced antimicrobial activity against all tested *M. abscessus* complex strains (MIC 16–64 µg/mL), as well as *M. bovis* BCG (MIC 32 µg/mL) and *M. tuberculosis* H37Ra (MIC 16 µg/mL). Furthermore, compound bearing 4-F substituent and methyl group (**19a**) showed good antimicrobial activity against *M. abscessus* complex strains, as well as *M. bovis* BCG and *M. tuberculosis* H37Ra (MIC 4–8 µg/mL respectively). However, other S-alkyl groups—ethyl (**19b**) and propyl (**19c**) in imidazole bearing 4-fluorophenyl substituent completely diminished antimicrobial activity against tested strains (Table 1).

#### 4. Conclusions

During this study, a series of imidazole-2-thiol bearing benzenesulfonamides was synthesized. To reach higher lipophilicity properties and potentially increase their membrane permeability through multidrug-resistant mycobacteria, various S-alkylation reactions were performed with alkyl halides.

Synthesized compounds showed structure-dependent antimicrobial activity against *Mycobacterium abscessus* complex strains. Furthermore, compounds **2–22a–c** showed little activity against multidrug-resistant Gram-positive and Gram-negative bacterial strains or drug-resistant fungi. However, 3-(2-thioxo-4-(4-(trifluoromethyl)phenyl)-2,3-dihydro-1H-imidazol-1-yl)benzenesulfonamide (**13**) has demonstrated high antibacterial activity against all tested mycobacterial strains and was more active than widely used antibiotics like rifampin, amikacin, or levofloxacin.

Previous studies have explored the impact of alkyl substitution on the antimicrobial activity of various compounds against mycobacteria and other clinically important pathogens. Oh et al. [39] have reported the synthesis of a series of novel *N*-Alkyl-5-hydroxypyrimidinone carboxamides as potent inhibitors of *M. tuberculosis* decaprenylphosphoryl-β-d-ribose 2'-oxidase. Faria et al. [40] describes alkyl promising activity and the high reactivity of alkyl hydrazide derivatives of isoniazid, suggesting that the alkylation is an important modification leading to the in vitro and in silico activity. Yang Yong et al. [41] described the synthesis of novel 8-alkylberberine derivatives bearing aliphatic chains and evaluated their antimicrobial activity. The study showed that increasing the length of the aliphatic chain had a significant

effect on the antibacterial activity of the compounds. However, antimicrobial activity started to decrease when alkyl chain consisted eight or more carbon atoms.

S-alkylation is widely employed strategy to increase the stability of biologically active compounds due to higher bond dissociation energy of the S-C bond compared to the N-C bond [42,43]. S-alkylated compounds are generally less susceptible to hydrolysis and more resistant to metabolic degradation compared to N-alkylated compounds, making S-alkylation an attractive strategy to enhance the biological activity of various compounds.

In our study, we compared S-alkylated benzenesulfonamide bearing imidazole derivatives against multidrug-resistant *M. abscessus* complex strains, and we found that 3-(4-(4-fluorophenyl)-2-(methylthio)-1H-imidazol-1-yl)benzenesulfonamide (**19a**) and 3-(4-(4-chlorophenyl)-2-(propylthio)-1H-imidazol-1-yl)benzenesulfonamide **18c** showed the highest antimycobacterial activity. For instance, MICs of compound **19a** with 4-fluorophenyl and S-methyl substituents against *M. abscessus* complex strains, as well as *M. bovis* BCG and *M. tuberculosis* H37Ra, were 4–8 µg/mL, respectively. However, ethyl or propyl groups in the same 1H-imidazol-2-thiol scaffold with 4-fluorophenyl group (compounds **19b** and **19c**) reduced the potency significantly. As for the imidazole scaffold with 4-chlorophenyl substituent, antimicrobial activity was increased by extending the alkyl chain. Compound **18c** containing S-propyl group was more potent than **18a** (S-methyl) and **18b** (S-ethyl) compounds.

These results suggest that the S-alkylated benzenesulfonamide-bearing imidazole derivatives could be further explored as a scaffold for the development of novel, multidrug-resistant *M. abscessus* complex-directed antimicrobials.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/microorganisms11040935/s1>, Figure S1: <sup>1</sup>H NMR of compound **3** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S2: <sup>13</sup>C NMR of compound **3** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S3: <sup>1</sup>H NMR of compound **6** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S4: <sup>13</sup>C NMR of compound **6** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S5: <sup>1</sup>H NMR of compound **10** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S6: <sup>13</sup>C NMR of compound **10** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S7: <sup>1</sup>H NMR of compound **13** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S8: <sup>13</sup>C NMR of compound **13** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S9: <sup>1</sup>H NMR of compound **16a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S10: <sup>13</sup>C NMR of compound **16a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S11: <sup>1</sup>H NMR of compound **16b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S12: <sup>13</sup>C NMR of compound **16b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S13: <sup>1</sup>H NMR of compound **16c** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S14: <sup>13</sup>C NMR of compound **16c** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S15: <sup>1</sup>H NMR of compound **17a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S16: <sup>13</sup>C NMR of compound **17a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S17: <sup>1</sup>H NMR of compound **17b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S18: <sup>13</sup>C NMR of compound **17b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S19: <sup>1</sup>H NMR of compound **18a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S20: <sup>13</sup>C NMR of compound **18a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S21: <sup>1</sup>H NMR of compound **18b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S22: <sup>13</sup>C NMR of compound **18b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S23: <sup>1</sup>H NMR of compound **18c** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S24: <sup>13</sup>C NMR of compound **18c** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S25: <sup>1</sup>H NMR of compound **19a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S26: <sup>13</sup>C NMR of compound **19a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S27: <sup>1</sup>H NMR of compound **19b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S28: <sup>13</sup>C NMR of compound **19b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S29: <sup>1</sup>H NMR of compound **19c** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S30: <sup>13</sup>C NMR of compound **19c** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S31: <sup>1</sup>H NMR of compound **20a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S32: <sup>13</sup>C NMR of compound **20a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S33: <sup>1</sup>H NMR of compound **20b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S34: <sup>13</sup>C NMR of compound **20b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S35: <sup>1</sup>H NMR of compound **20c** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S36: <sup>13</sup>C NMR of compound **20c** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S37: <sup>1</sup>H NMR of compound **21a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S38: <sup>13</sup>C NMR of compound **21a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S39: <sup>1</sup>H NMR of compound **21b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S40: <sup>13</sup>C NMR of compound **21b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S41: <sup>1</sup>H NMR of compound **21c** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S42: <sup>13</sup>C NMR of compound **21c** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S43: <sup>1</sup>H NMR of compound **22a** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S44: <sup>13</sup>C NMR of compound **22a** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S45: <sup>1</sup>H NMR of compound **22b** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S46: <sup>13</sup>C NMR of compound **22b** at 101 MHz (DMSO-*d*<sub>6</sub>), Figure S47: <sup>1</sup>H NMR of compound **22c** at 400 MHz (DMSO-*d*<sub>6</sub>), Figure S48: <sup>13</sup>C NMR of compound **22c** at 101

MHz (DMSO-*d*<sub>6</sub>). Table S1: The in vitro antibacterial activity of compounds **2-22c** against multidrug-resistant bacterial strains; Table S2: The in vitro antifungal activity of compounds **2-22c** against multidrug-resistant *Candida auris* strains.

**Author Contributions:** Conceptualization, V.M. and P.K.; methodology, V.M., B.B. and P.K.; synthesis, B.B.; investigation, B.B., B.G., R.P., V.P., A.G., E.N., R.G. and P.K.; writing—original draft preparation, B.B. and P.K.; writing—review and editing, V.M. and P.K.; supervision, V.M. and P.K. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was funded by The Doctoral Fund of Kaunas University of Technology No. A-410, approved 26 June 2019, Kaunas, Lithuania.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Data is contained within the article and supplementary materials. The compounds are available from the corresponding author.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Abdelaal, H.F.M.; Chan, E.D.; Young, L.; Baldwin, S.L.; Coler, R.N. Mycobacterium Abscessus: It's Complex. *Microorganisms* **2022**, *10*, 1454. [[CrossRef](#)]
2. Shiraishi, K.; Kasai, H.; Saito, M.; Kawaguchi, H.; Kinoshita, T.; Suzuki, T.; Shikano, K.; Takagi, K.; Sakao, S.; Hanazawa, T.; et al. Case of a Deep Neck Abscess During Treatment for COVID-19. *Am. J. Case Rep.* **2022**, *23*, e936034-1. [[CrossRef](#)] [[PubMed](#)]
3. Furuuchi, K.; Morimoto, K.; Yoshiyama, T.; Tanaka, Y.; Fujiwara, K.; Okumura, M.; Izumi, K.; Shiraishi, Y.; Mitarai, S.; Ogata, H.; et al. Interrelational Changes in the Epidemiology and Clinical Features of Nontuberculous Mycobacterial Pulmonary Disease and Tuberculosis in a Referral Hospital in Japan. *Respir. Med.* **2019**, *152*, 74–80. [[CrossRef](#)]
4. Shin, S.H.; Jhun, B.W.; Kim, S.-Y.; Choe, J.; Jeon, K.; Huh, H.J.; Ki, C.-S.; Lee, N.Y.; Shin, S.J.; Daley, C.L.; et al. Nontuberculous Mycobacterial Lung Diseases Caused by Mixed Infection with Mycobacterium Avium Complex and Mycobacterium Abscessus Complex. *Antimicrob. Agents Chemother.* **2018**, *62*, e01105-18. [[CrossRef](#)] [[PubMed](#)]
5. Philley, J.V.; Griffith, D.E. Medical Management of Pulmonary Nontuberculous Mycobacterial Disease. *Thorac. Surg. Clin.* **2019**, *29*, 65–76. [[CrossRef](#)] [[PubMed](#)]
6. Yan, Q.; Wang, W.; Zhao, W.; Zuo, L.; Wang, D.; Chai, X.; Cui, J. Differentiating Nontuberculous Mycobacterium Pulmonary Disease from Pulmonary Tuberculosis through the Analysis of the Cavity Features in CT Images Using Radiomics. *BMC Pulm. Med.* **2022**, *22*, 4. [[CrossRef](#)] [[PubMed](#)]
7. Kumar, K.; Daley, C.L.; Griffith, D.E.; Loebinger, M.R. Management of Mycobacterium Avium Complex and Mycobacterium Abscessus Pulmonary Disease: Therapeutic Advances and Emerging Treatments. *Eur. Respir. Rev.* **2022**, *31*, 210212. [[CrossRef](#)]
8. Togo, T.; Atsumi, J.; Hiramatsu, M.; Shimoda, K.; Morimoto, K.; Shiraishi, Y. Outcomes of Surgical Treatment for Mycobacterium Abscessus Complex Pulmonary Disease. *Ann. Thorac. Surg.* **2022**, *113*, 949–956. [[CrossRef](#)]
9. Qin, R.; Wang, P.; Wang, B.; Fu, L.; Batt, S.M.; Besra, G.S.; Wu, C.; Wang, Y.; Huang, H.; Lu, Y.; et al. Identification of Thiophene-Benzenesulfonamide Derivatives for the Treatment of Multidrug-Resistant Tuberculosis. *Eur. J. Med. Chem.* **2022**, *231*, 114145. [[CrossRef](#)]
10. Du, J.; Liu, P.; Zhu, Y.; Wang, G.; Xing, S.; Liu, T.; Xia, J.; Dong, S.; Lv, N.; Li, Z. Novel Tryptanthrin Derivatives with Benzenesulfonamide Substituents: Design, Synthesis, and Anti-Inflammatory Evaluation. *Eur. J. Med. Chem.* **2023**, *246*, 114956. [[CrossRef](#)]
11. Audat, S.A.; Al-Shar'i, N.A.; Al-Oudat, B.A.; Alnabulsi, S. Design, Synthesis, and Biological Evaluation of SMYD3 Inhibitors Possessing N-Thiazole Benzenesulfonamide Moiety as Potential Anti-Cancer Agents. *J. Saudi Chem. Soc.* **2022**, *26*, 101482. [[CrossRef](#)]
12. Muthukumar, R.; Karnan, M.; Elangovan, N.; Karunanidhi, M.; Thomas, R. Synthesis, Spectral Analysis, Antibacterial Activity, Quantum Chemical Studies and Supporting Molecular Docking of Schiff Base (E)-4-((4-Bromobenzylidene) Amino)Benzenesulfonamide. *J. Indian Chem. Soc.* **2022**, *99*, 100405. [[CrossRef](#)]
13. Dhumad, A.M.; Jassem, A.M.; Alharis, R.A.; Almashal, F.A. Design, Cytotoxic Effects on Breast Cancer Cell Line (MDA-MB 231), and Molecular Docking of Some Maleimide-Benzenesulfonamide Derivatives. *J. Indian Chem. Soc.* **2021**, *98*, 100055. [[CrossRef](#)]
14. Kanagavalli, A.; Jayachitra, R.; Thilagavathi, G.; Padmavathy, M.; Elangovan, N.; Sowrirajan, S.; Thomas, R. Synthesis, Structural, Spectral, Computational, Docking and Biological Activities of Schiff Base (E)-4-Bromo-2-Hydroxybenzylidene) Amino)-N-(Pyrimidin-2-Yl) Benzenesulfonamide from 5-Bromosalicylaldehyde and Sulfadiazine. *J. Indian Chem. Soc.* **2023**, *100*, 100823. [[CrossRef](#)]
15. Carrión, M.D.; Rubio-Ruiz, B.; Franco-Montalban, F.; Amoia, P.; Zuccarini, M.C.; De Simone, C.; Camacho, M.E.; Amoroso, R.; Maccallini, C. New Amidine-Benzenesulfonamides as INOS Inhibitors for the Therapy of the Triple Negative Breast Cancer. *Eur. J. Med. Chem.* **2023**, *248*, 115112. [[CrossRef](#)] [[PubMed](#)]



16. El-Azab, A.S.; Alkahtani, H.M.; AlSaif, N.A.; Al-Suwaidan, I.A.; Obaidullah, A.J.; Alanazi, M.M.; Al-Obaid, A.M.; Al-Agamy, M.H.M.; Abdel-Aziz, A.A.-M. Synthesis, Antiproliferative and Enzymatic Inhibition Activities of Quinazolines Incorporating Benzenesulfonamide: Cell Cycle Analysis and Molecular Modeling Study. *J. Mol. Struct.* **2023**, *1278*, 134928. [[CrossRef](#)]
17. Balandis, B.; Šimkūnas, T.; Paketurytė-Latvė, V.; Michailovienė, V.; Mickevičiūtė, A.; Manakova, E.; Gražulis, S.; Belyakov, S.; Kairys, V.; Mickevičius, V.; et al. Beta and Gamma Amino Acid-Substituted Benzenesulfonamides as Inhibitors of Human Carbonic Anhydrases. *Pharmaceuticals* **2022**, *15*, 477. [[CrossRef](#)]
18. Kakakhan, C.; Türkeş, C.; Güleç, Ö.; Demir, Y.; Arslan, M.; Özkemahlı, G.; Beydemir, Ş. Exploration of 1,2,3-Triazole Linked Benzenesulfonamide Derivatives as Isoform Selective Inhibitors of Human Carbonic Anhydrase. *Bioorganic Med. Chem.* **2023**, *77*, 117111. [[CrossRef](#)]
19. Vaškevičienė, I.; Paketurytė, V.; Zubrienė, A.; Kantminienė, K.; Mickevičius, V.; Matulis, D. N-Sulfamoylphenyl- and N-Sulfamoylphenyl-N-Thiazolyl-β-Alanines and Their Derivatives as Inhibitors of Human Carbonic Anhydrases. *Bioorganic Chem.* **2017**, *75*, 16–29. [[CrossRef](#)]
20. Aspatwar, A.; Winum, J.-Y.; Carta, F.; Supuran, C.T.; Hammaren, M.; Parikka, M.; Parkkila, S. Carbonic Anhydrase Inhibitors as Novel Drugs against Mycobacterial β-Carbonic Anhydrases: An Update on In Vitro and In Vivo Studies. *Molecules* **2018**, *23*, 2911. [[CrossRef](#)]
21. Stahl, D.A.; Urbance, J.W. The Division between Fast- and Slow-Growing Species Corresponds to Natural Relationships among the Mycobacteria. *J. Bacteriol.* **1990**, *172*, 116–124. [[CrossRef](#)]
22. Aspatwar, A.; Hammarén, M.; Koskinen, S.; Luukinen, B.; Barker, H.; Carta, F.; Supuran, C.T.; Parikka, M.; Parkkila, S. β-CA-Specific Inhibitor Dithiocarbamate Fc14–584B: A Novel Antimycobacterial Agent with Potential to Treat Drug-Resistant Tuberculosis. *J. Enzym. Inhib. Med. Chem.* **2017**, *32*, 832–840. [[CrossRef](#)] [[PubMed](#)]
23. Nishimori, I.; Minakuchi, T.; Vullo, D.; Scozzafava, A.; Innocenti, A.; Supuran, C.T. Carbonic Anhydrase Inhibitors. Cloning, Characterization, and Inhibition Studies of a New β-Carbonic Anhydrase from Mycobacterium Tuberculosis. *J. Med. Chem.* **2009**, *52*, 3116–3120. [[CrossRef](#)]
24. Muhammed, M.T.; Er, M.; Akkoc, S. Molecular Modeling and in Vitro Antiproliferative Activity Studies of Some Imidazole and Isoxazole Derivatives. *J. Mol. Struct.* **2023**, *1282*, 135066. [[CrossRef](#)]
25. Çetiner, G.; Acar Çevik, U.; Celik, I.; Bostancı, H.E.; Özkay, Y.; Kaplancıklı, Z.A. New Imidazole Derivatives as Aromatase Inhibitor: Design, Synthesis, Biological Activity, Molecular Docking, and Computational ADME-Tox Studies. *J. Mol. Struct.* **2023**, *1278*, 134920. [[CrossRef](#)]
26. Roy, D.; Anas, M.; Manhas, A.; Saha, S.; Kumar, N.; Panda, G. Synthesis, Biological Evaluation, Structure—Activity Relationship Studies of Quinoline-Imidazole Derivatives as Potent Antimalarial Agents. *Bioorganic Chem.* **2022**, *121*, 105671. [[CrossRef](#)] [[PubMed](#)]
27. Liu, Y.-C.; Yang, Y.-D.; Liu, W.-Q.; Du, T.-T.; Wang, R.; Ji, M.; Yang, B.-B.; Li, L.; Chen, X.-G. Benzobis(Imidazole) Derivatives as STAT3 Signal Inhibitors with Antitumor Activity. *Bioorganic Med. Chem.* **2022**, *65*, 116757. [[CrossRef](#)] [[PubMed](#)]
28. Badura, A.; Krysiński, J.; Nowaczyk, A.; Buciuński, A. Application of Artificial Neural Networks to the Prediction of Antifungal Activity of Imidazole Derivatives against *Candida Albicans*. *Chemom. Intell. Lab. Syst.* **2022**, *222*, 104501. [[CrossRef](#)]
29. Mickevičienė, K.; Voskienė, A.; Mickevičius, V. Synthesis of Some 1- and 2-Carboxyalkyl Substituted Benzimidazoles and Their Derivatives. *Res. Chem. Intermed.* **2014**, *40*, 1619–1631. [[CrossRef](#)]
30. Tumosienė, I.; Peleckis, A.; Jonuškienė, I.; Vaickelionienė, R.; Kantminienė, K.; Šiugždaitė, J.; Beresnevičius, Z.J.; Mickevičius, V. Synthesis of Novel 1,2- and 2-Substituted Benzimidazoles with High Antibacterial and Antioxidant Activity. *Monatsh. Chem.* **2018**, *149*, 577–594. [[CrossRef](#)]
31. Strelciunaite, V.; Jonuskiene, I.; Anusevicius, K.; Tumsiene, I.; Siugzdaite, J.; Ramanauskaite, I.; Mickevicius, V. Synthesis of Novel Benzimidazoles 2-Functionalized with Pyrrolidinone and γ-Amino Acid with a High Antibacterial Activity. *Heterocycles* **2016**, *92*, 235. [[CrossRef](#)]
32. Kuzu, B.; Tan, M.; Taslimi, P.; Gülçin, İ.; Taşpınar, M.; Menges, N. Mono- or Di-Substituted Imidazole Derivatives for Inhibition of Acetylcholine and Butyrylcholine Esterases. *Bioorganic Chem.* **2019**, *86*, 187–196. [[CrossRef](#)]
33. Perevalov, V.P.; Mityanov, V.S.; Lichitsky, B.V.; Komogortsev, A.N.; Kuz'mina, L.G.; Koldaeva, T.Y.; Miroshnikov, V.S.; Kutasevich, A.V. Synthesis of Highly Functional Imidazole Derivatives via Assembly of 2-Unsubstituted Imidazole N-Oxides with CH-Acids and Arylglyoxals. *Tetrahedron* **2020**, *76*, 130947. [[CrossRef](#)]
34. Cirillo, D.; Angelucci, F.; Bjørsvik, H.-R. Functionalization of the Imidazole Backbone by Means of a Tailored and Optimized Oxidative Heck Cross-Coupling. *Adv. Synth. Catal.* **2020**, *362*, 5079–5092. [[CrossRef](#)]
35. Balandis, B.; Mickevičius, V.; Petrikaitė, V. Exploration of Benzenesulfonamide-Bearing Imidazole Derivatives Activity in Triple-Negative Breast Cancer and Melanoma 2D and 3D Cell Cultures. *Pharmaceuticals* **2021**, *14*, 1158. [[CrossRef](#)] [[PubMed](#)]
36. Pfaller, M.A.; Andes, D.R.; Diekema, D.J.; Horn, D.L.; Reboli, A.C.; Rotstein, C.; Franks, B.; Azie, N.E. Epidemiology and Outcomes of Invasive Candidiasis Due to Non-*Albicans* Species of *Candida* in 2,496 Patients: Data from the Prospective Antifungal Therapy (PATH) Registry 2004–2008. *PLoS ONE* **2014**, *9*, e101510. [[CrossRef](#)]
37. Peano, A.; Beccati, M.; Chiavassa, E.; Pasquetti, M. Evaluation of the Antifungal Susceptibility of *Malassezia Pachydermatis* to Clotrimazole, Miconazole and Thiabendazole Using a Modified CLSI M27-A3 Microdilution Method. *Vet. Dermatol.* **2012**, *23*, 131-e29. [[CrossRef](#)]

38. Novak, I.; Klasinc, L.; McGlynn, S.P. Electronic Structure and Tautomerism of Thioamides. *J. Electron Spectrosc. Relat. Phenom.* **2016**, *209*, 62–65. [[CrossRef](#)]
39. Oh, S.; Park, Y.; Engelhart, C.A.; Wallach, J.B.; Schnappinger, D.; Arora, K.; Manikkam, M.; Gac, B.; Wang, H.; Murgolo, N.; et al. Discovery and Structure–Activity-Relationship Study of N-Alkyl-5-Hydroxypyrimidinone Carboxamides as Novel Antitubercular Agents Targeting Decaprenylphosphoryl- $\beta$ -d-Ribose 2'-Oxidase. *J. Med. Chem.* **2018**, *61*, 9952–9965. [[CrossRef](#)]
40. de Faria, C.F.; Moreira, T.; Lopes, P.; Costa, H.; Krewall, J.R.; Barton, C.M.; Santos, S.; Goodwin, D.; Machado, D.; Viveiros, M.; et al. Designing New Antitubercular Isoniazid Derivatives with Improved Reactivity and Membrane Trafficking Abilities. *Biomed. Pharmacother.* **2021**, *144*, 112362. [[CrossRef](#)]
41. Yong, Y.; Xiao-li, Y.; Xue-gang, L.; Jing, Z.; Baoshun, Z.; Lujiang, Y. Synthesis and Antimicrobial Activity of 8-Alkylberberine Derivatives with a Long Aliphatic Chain. *Planta Med.* **2007**, *73*, 602–604. [[CrossRef](#)] [[PubMed](#)]
42. Hagar, M.; Soliman, S.M.; Ibid, F.; El Ashry, E.S.H. Synthesis, Molecular Structure and Spectroscopic Studies of Some New Quinazolin-4(3H)-One Derivatives; an Account on the N- versus S-Alkylation. *J. Mol. Struct.* **2016**, *1108*, 667–679. [[CrossRef](#)]
43. Spears, R.J.; McMahon, C.; Chudasama, V. Cysteine Protecting Groups: Applications in Peptide and Protein Science. *Chem. Soc. Rev.* **2021**, *50*, 11098–11155. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.